

Committee

Microscopy and Imaging Sub-committee

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Scientist – II (Microscopy and Imaging – CCRF)

Booking

General Instructions

General Instructions for Microscopy and Imaging Sub-Facility, CCRF Users:

Objectives for use:

- a. 3D and 4D image acquisition
- b. Multi-time series live cell-imaging (with control of gas environment and moisture around the culture plate/slide etc.)
- c. FRAP
- d. FRET
- e. Photo activation and conversion
- f. Co-localization of 2-5 signals. (NOT Double or triple labelling that can be observed and imaged on a regular fluorescent imaging microscope)
- g. Spectral analysis for:
 - i. Spectral imaging and unmixing
 - ii. Auto-fluorescence separation by online spectral un-mixing
 - iii. Histogram analysis of emission spectrum
- h. Obtaining z-stacks
- i. 3D analysis to evaluate & display 3D image data stacks with measuring tools
- j. 3D visualization & multichannel volume rendering of 3D stacks

- k. Image reconstruction
- l. Measurements across z-stack
- m. Movie co-localization with intensity profiles for quantification
- n. Real time ratio-display
- o. 2D and 3D image deconvolution

Method:

1. Users are required to check the CCRF website and navigate to the Microscopy and Imaging sub-facility tab that has the booking form:
The following sample details need to be provided:
 - i. Sample type
 - ii. Purpose
 - iii. Type of imaging/experiment
 - iv. Required time slot
 - v. Other equipment required- gas cylinders, cell incubator, centrifuge, refrigerator, live imaging setup
2. Sample preparation guidelines are mentioned below, and users must strictly adhere to these guidelines:
 - i. Only confocal grade glass slides, cover slips, plates and dishes are to be used; cover slips mandatorily must be of 1 or 1.5 number with approximately 0.17 mm thickness.
 - ii. In case one is not sure about the glass slides, coverslips, etc. please consult the scientist before you perform the experiment.
 - iii. Users must be fully aware of the excitation and emission wavelengths of the fluorophores/ dyes that they have used in their experiments.
 - iv. It is the responsibility of the user to collect the data after acquiring the images. Data in the main system is periodically deleted to free up the system space. The facility will not be responsible for any loss of data.
3. The scientist in-charge should be consulted before the experiment is planned to discuss the reagents, experimental paradigms, sample size, etc. for the planned experiment.
4. Consultation for experiment (before booking) can be done verbally by visiting the facility, through mobile/ telephone or via e-mail.
5. The user should bring all the reagents required (The facility has basic requirements like gas cylinders, refrigerator, and stage-incubators for incubation. Basic dyes like toluidine blue, methylene blue, cell-counting chamber, shakers, etc. are to be provided by the user; the user can get in touch with the General Facility, CCRF to see if these reagents/ equipment are available).
6. The booking window is open 45 days and 30 days in advance for the confocal/ airyscan and the image processing system respectively; it is

the responsibility of the users to plan their experiments and book the slots ahead of time.

7. At least one-week prior notice should be given for cancellation, so that the slot may be offered to another person; otherwise, charges will be levied.
8. Analysis is to be conducted on the dedicated offline system, so that the online system may be kept free for users. Data will be provided only in formatted hard drives. No pen drives should be used on the system unless prior formatting is done in view of/by the scientist/ technical officer.
9. User logs are maintained both electronically and as hard copy. Users should sign the register once their experiments are complete and they have received their data.

Charges

Confocal/ Airyscan Scanning (Slides/ Multi-well plates): ₹200/- per 1.5 hours

Live-cell Imaging (Using Stage Incubator): ₹500/- for the first hour and subsequently ₹100/- per hour if experiment is run uninterrupted

Image Processing: ₹100/- per 2 hours

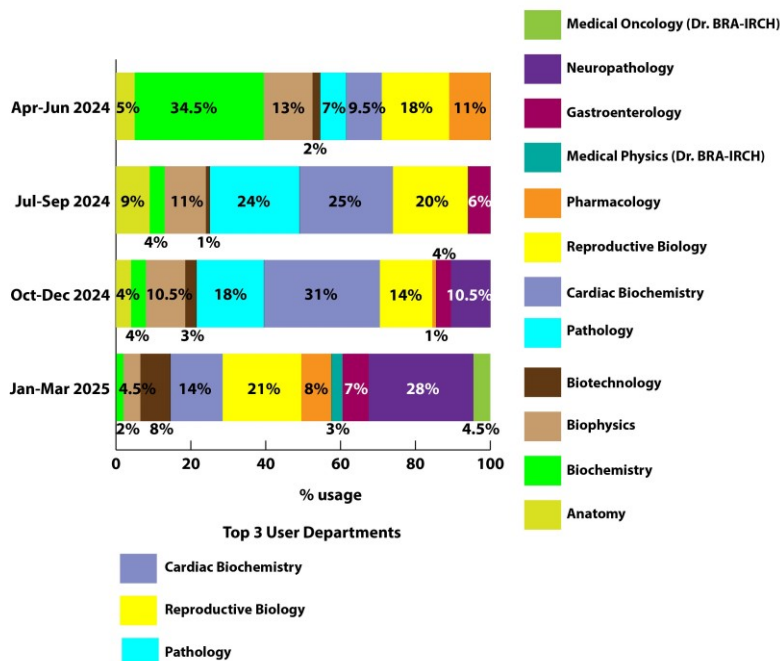
Usage Graphs

Department-wise usage of LSM980 Confocal Microscope in 2025-2026



Usage Statistics for the Financial Year 2025-2026

Department-wise usage of LSM980 Confocal Microscope in 2024-2025



Usage Statistics for the Financial Year 2024-2025

Trainings/ Workshops

Upcoming Workshops

We conduct 1-2 days training workshops in the spring (tentatively March/ April) and fall (tentatively September/ October) each year. Please look out for the calls through emails and community groups.

Past Workshops

Summer Confocal Microscopy & Image Analysis Hands-on Training Workshop was held on April 1st and 2nd, 2026

Fall Confocal Microscopy & Image Analysis Lectures cum Hands-on Training Workshop was held on September 1st and 2nd, 2025

Summer Image Analysis Hands-on Training Workshop was held on June 12th and 13th, 2025

Advanced 3D Image Processing Workshop using Arivis was held on January 21st, 2025

Fall Live-Cell Imaging Hands-on Training Workshop cum Lecture was held on September 4th and 5th, 2024

Spring Confocal and Super-Resolution Microscopy Hands-on Training Workshop was held on April 19th and 20th, 2024

Confocal Microscopy Lecture cum Hands-on Training Workshop was held on October 11th, 2023

Webinar on Confocal Microscopy, Specifications and Applications of the Zeiss LSM980 System was held on March 8th, 2021

Demonstration of Zeiss LSM980 Confocal system installed in CCRF was held on March 4th, 2021

Contact Us

Confocal Microscopy Facility - CCRF

Room No. 8A, Convergence Block, Ground Floor

(along the Department of Anatomy hallway)

For queries, sample preparation and bookings email us at:

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Events Photo Gallery



Prof. Randeep Guleria (Director, AIIMS), Prof. Subrata Sinha (Dean, Research) and Prof. Parthaprasad Chattopadhyay (Faculty Coordinator, Microscopy and Imaging sub-facility, CCRF) addressing the audience during the inauguration of the sub-facility on March 8th, 2021.



Dr. Tony George Jacob (Member, Microscopy and Imaging sub-facility, CCRF) accompanying Prof. Randeep Guleria (Director, AIIMS) to the SET Facility Studio on the inauguration of the sub-facility on March 8th, 2021.



Prof. Randeep Guleria (Director, AIIMS), Prof. Subrata Sinha (Dean, Research), Prof. Vineet Ahuja (Associate Dean, Research) and Prof. Chitra Sarkar (Former Dean, Research) having a discussion on the sidelines of the inauguration of the sub-facility on March 8th, 2021.



Dr. Saumitra Dey Choudhury (Scientist-II, CCRF) and Dr. Somenath Ghatak (Application Specialist, Carl Zeiss India) during the Demonstration of Zeiss LSM980 Confocal system installed in CCRF on March 4th, 2021.



Dr. Manoj Manna (Products and Application Specialist, Leica Microsystems India) and Mr. Vishal Singh (Laboratory Technician, SAIF/ Anatomy) with trainees during the Confocal Microscopy Hands-on Training Workshop held on October 11th, 2023.



Dr. Somenath Ghatak (Application Specialist, Carl Zeiss India) with trainees during the Confocal Microscopy Hands-on Training Workshop held on October 11th, 2023.



Participants during the Advanced 3D Image Processing Workshop using Arivis held on January 21st, 2025.



Participants during the Summer Image Analysis Hands-on Training Workshop held on June 12th and 13th, 2025.



Participants during the Summer Confocal Microscopy and Image Analysis Hands-on Training Workshop held on April 1st and 2nd, 2026.